# **RAMAPO** COLLEGE OF NEW IERSEY

# Introduction

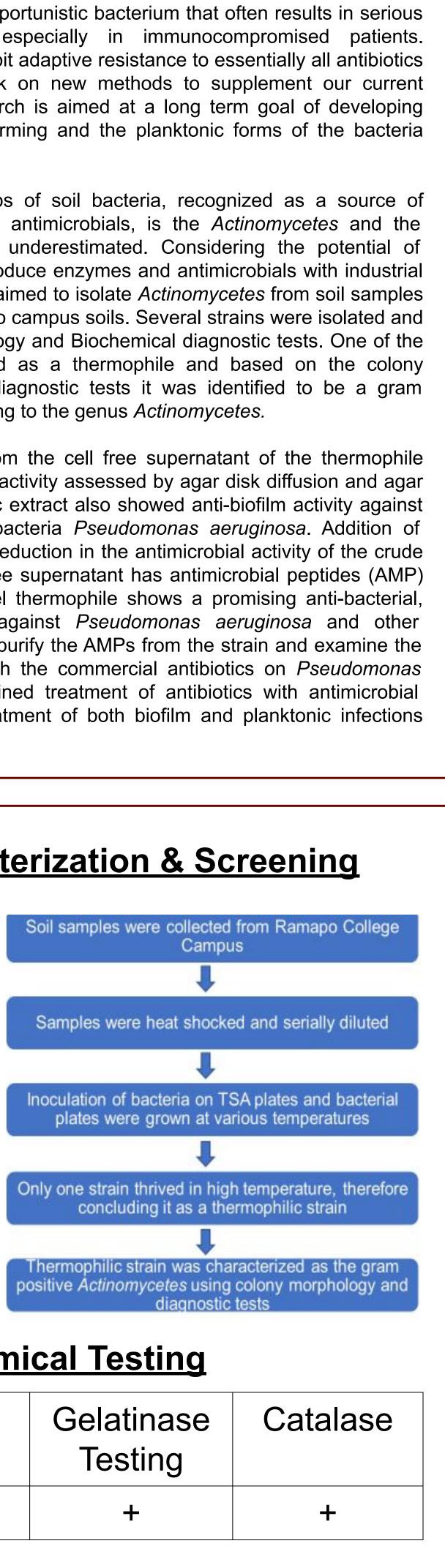
Pseudomonas aeruginosa is an opportunistic bacterium that often results in serious infections in health-care settings especially in immunocompromised patients. Pseudomonas growing in biofilms exhibit adaptive resistance to essentially all antibiotics and there is an urgent need to work on new methods to supplement our current antibiotics. Much of our current research is aimed at a long term goal of developing anti-biofilm agents targeting biofilm forming and the planktonic forms of the bacteria Pseudomonas aeruginosa.

One of the most important groups of soil bacteria, recognized as a source of commercially important enzymes and antimicrobials, is the Actinomycetes and the diversity of Actinomycetes is largely underestimated. Considering the potential of discovering Actinomycetes that can produce enzymes and antimicrobials with industrial and medical applications, this study is aimed to isolate *Actinomycetes* from soil samples collected from selected sites of Ramapo campus soils. Several strains were isolated and screened using the standard Microbiology and Biochemical diagnostic tests. One of the several strains isolated was identified as a thermophile and based on the colony morphology, gram staining and the diagnostic tests it was identified to be a gram positive spore forming bacteria belonging to the genus Actinomycetes.

The extracellular crude extract from the cell free supernatant of the thermophile exhibited anti-bacterial and anti-fungal activity assessed by agar disk diffusion and agar well diffusion method. The thermophilic extract also showed anti-biofilm activity against the bacterial biofilm formed by the bacteria Pseudomonas aeruginosa. Addition of Proteinase-K to the extract showed a reduction in the antimicrobial activity of the crude extract which indicated that the cell-free supernatant has antimicrobial peptides (AMP) inhibiting the test microbes. The novel thermophile shows a promising anti-bacterial, anti-fungal and anti-biofilm activity against Pseudomonas aeruginosa and other microbes. The future goal is to further purify the AMPs from the strain and examine the synergistic effects of the peptides with the commercial antibiotics on *Pseudomonas* aeruginosa biofilm inhibition. A combined treatment of antibiotics with antimicrobial peptides may offer a very potent treatment of both biofilm and planktonic infections resulting in novel adjuvant therapies.

# **Isolation, Characterization & Screening**

Ammonium sulphate at a 60% saturation (w/v) was used to precipitate the extracellular AMP from the crude cell-free extract. To further purify and remove the ultra-filtration ammonium sulphate. technique (using Amicon filters) was used. The concentration of the partially purified AMP was measured using the Bradford Assay. Using the disk diffusion method, the antimicrobial activity was measured using the zones of inhibition against the gram positive and gram negative bacteria, Staphylococcus aureus, Pseudomonas aeruginosa and Escherichia coli. The antifungal activity was measured against Candida albicans. The results showed that the extract had both anti-bacterial, anti-fungal and anti-biofilm activity against *Pseudomonas aeruginosa* biofilms.



# **Biochemical Testing**

Starch Hydrolysis	MR-VP	Gelatinase Testing
+	+	+



## Isolation, Identification and Assessment of the Antimicrobial Activities of a Thermophilic Actinomycetes Strain Extracted from the Soil. Dr. Suvekshya Kokila Kota, Juhi Desai, Shrestha, Mariam Tinawi

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