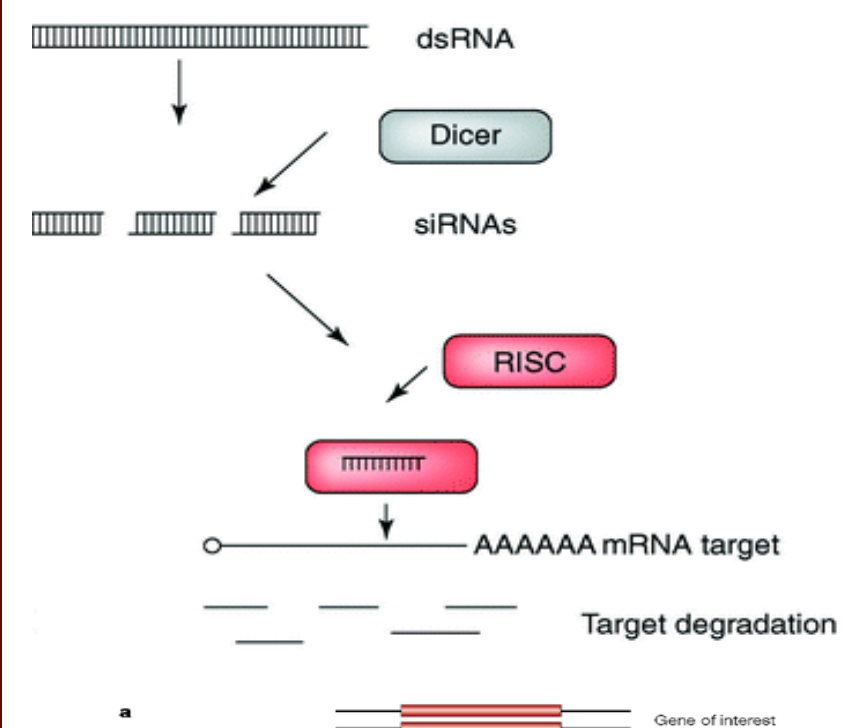


RNA-mediated interference (RNAi) is a process by which RNA molecules inhibit gene expression via specific degradation of mRNA transcripts. Since its discovery nearly 20 years ago, researchers have utilized this understanding to specifically knock-down genes of interest. In the nematode *C. elegans*, several RNAi techniques have been developed, including injection of double stranded RNA (dsRNA) and feeding of bacteria expressing dsRNA, yet a systematic study determining the efficacy of the two approaches has not been conducted to date. In this study, we test the efficacy of the injection and feeding method to knock-down an essential mitotic protein HCP-3, and assess the effectiveness of this strategy for use in future studies. Knock-down effectiveness will be measured by western blot analysis and immunofluorescence, and the phenotypic consequences will be assayed by embryonic lethality and live-imaging of cell division in mCherry:Histone-H2B & GFP:α-tubulin transgenic worms. Currently, HCP-3 RNAi feeding demonstrates that knock-downs result in 100% embryonic lethality and western blot analysis reveals that HCP-3 has been depleted to ~5% that of wild-type levels. We are currently in the process of testing the injection method and replicating the feeding results.

RNA-mediated interference (RNAi)

- inhibit gene expression by causing the destruction of specific mRNA molecules
- when mRNA molecules are no longer present, protein cannot be made



Mechanism of RNA-mediated Interference (RNAi)

- dsRNA recognized and chopped into siRNA pieces by the enzyme Dicer
- siRNAs bind to the RISC complex and find complementary mRNA
- RISC cleaves the mRNA transcript preventing translation & thus gene expression of the target transcript.

2 RNAi Strategies in *C. elegans*

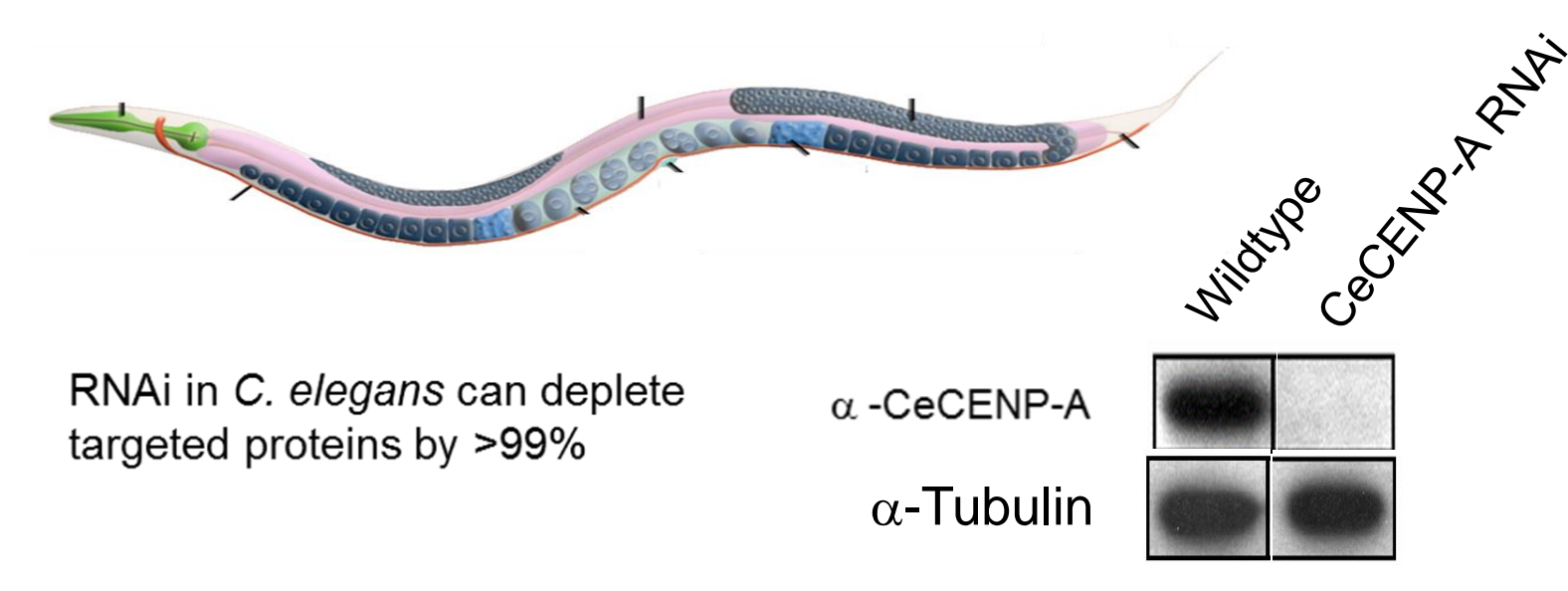
The gene of interest is inserted into a vector with 2 flanking Bacterial Promoters (T7).

- 1) transformed into *E. coli* -fed to the worms
- 2) transcribed into dsRNA -injected into the worms

C. elegans as a model for studying protein depletion

- Easy to maintain
- Hermaphrodites
- self-fertilize
- progeny are identical to parent
- High levels of gene knock-down via RNAi

As the CeCENP-A is depleted by the RNAi, the amount of protein in the cell decreases until there is nothing left

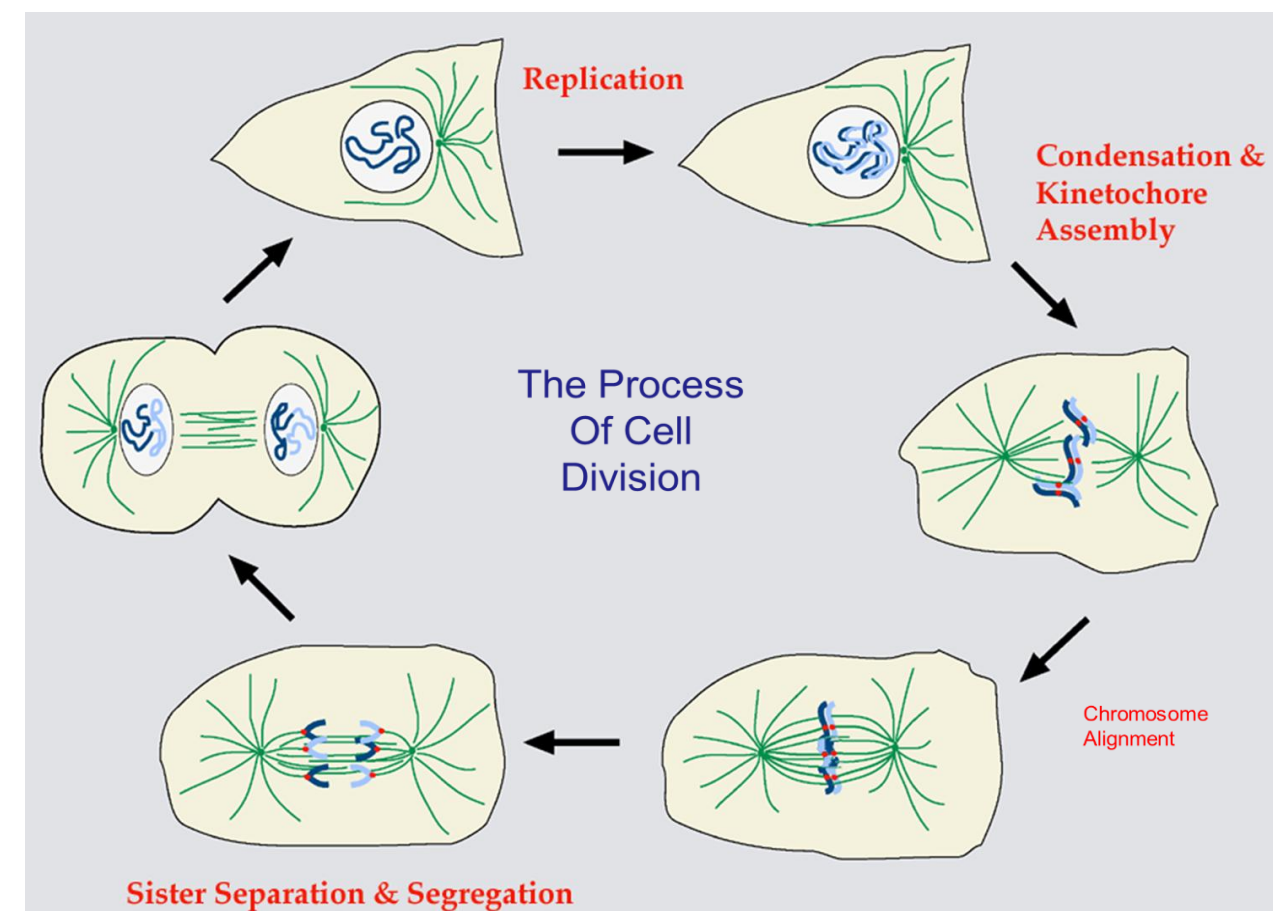
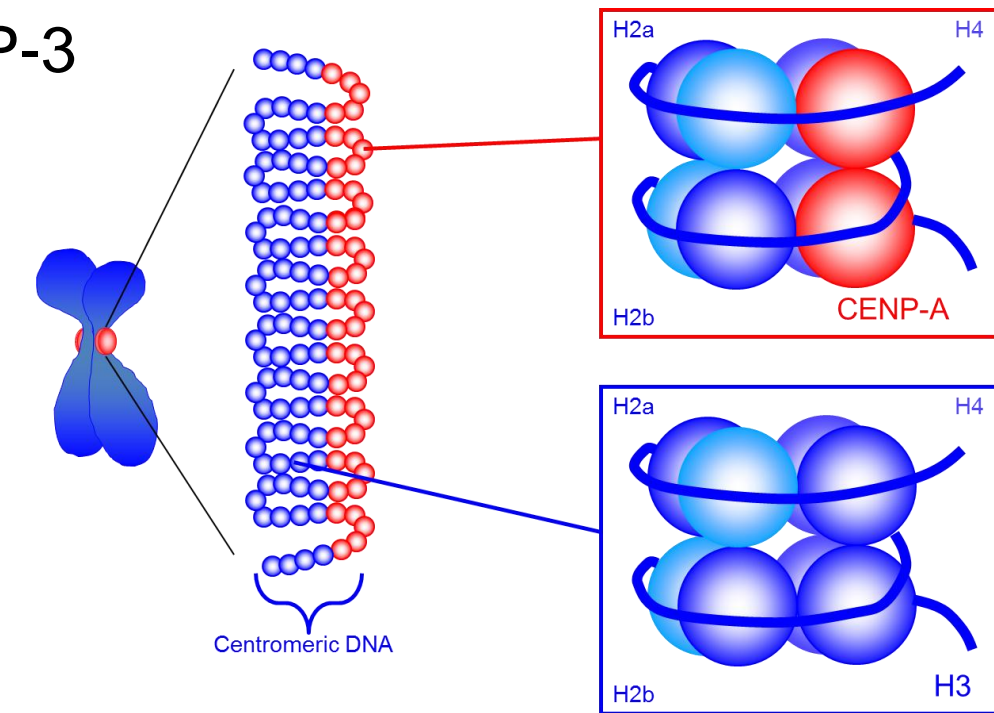


CENP-A's role in mitosis

-CENP-A is a protein located within the centromere of a chromosome & essential for chromosome segregation.

-In *C. elegans*, two homologs of CENP-A exist:

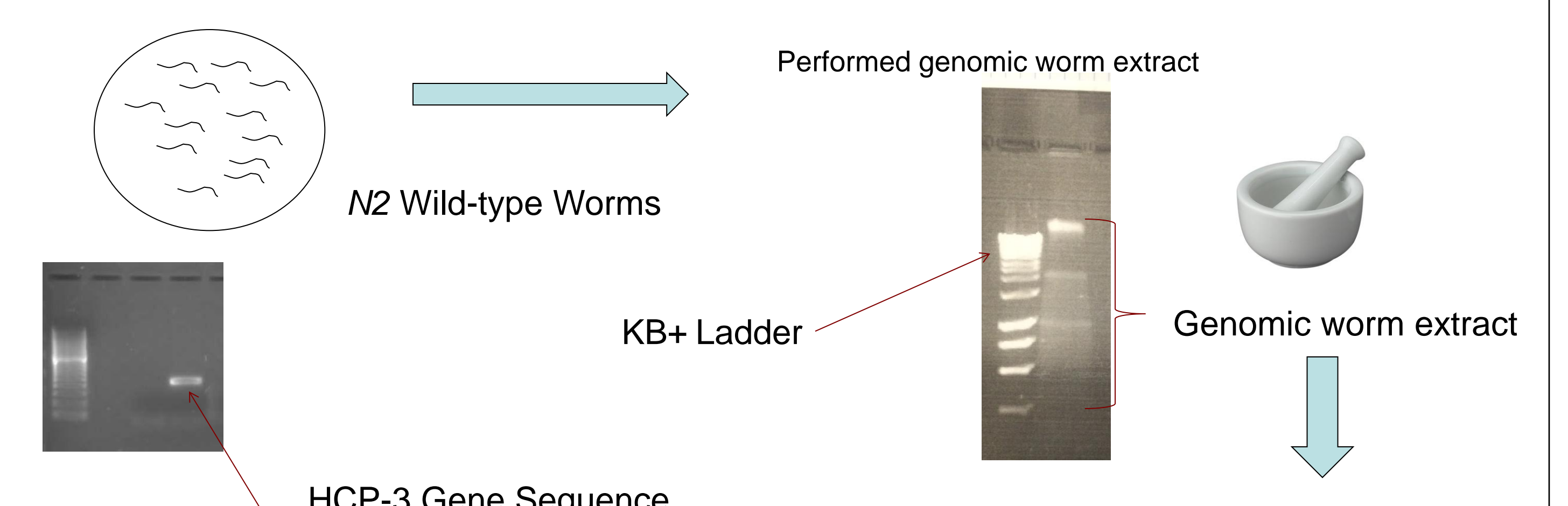
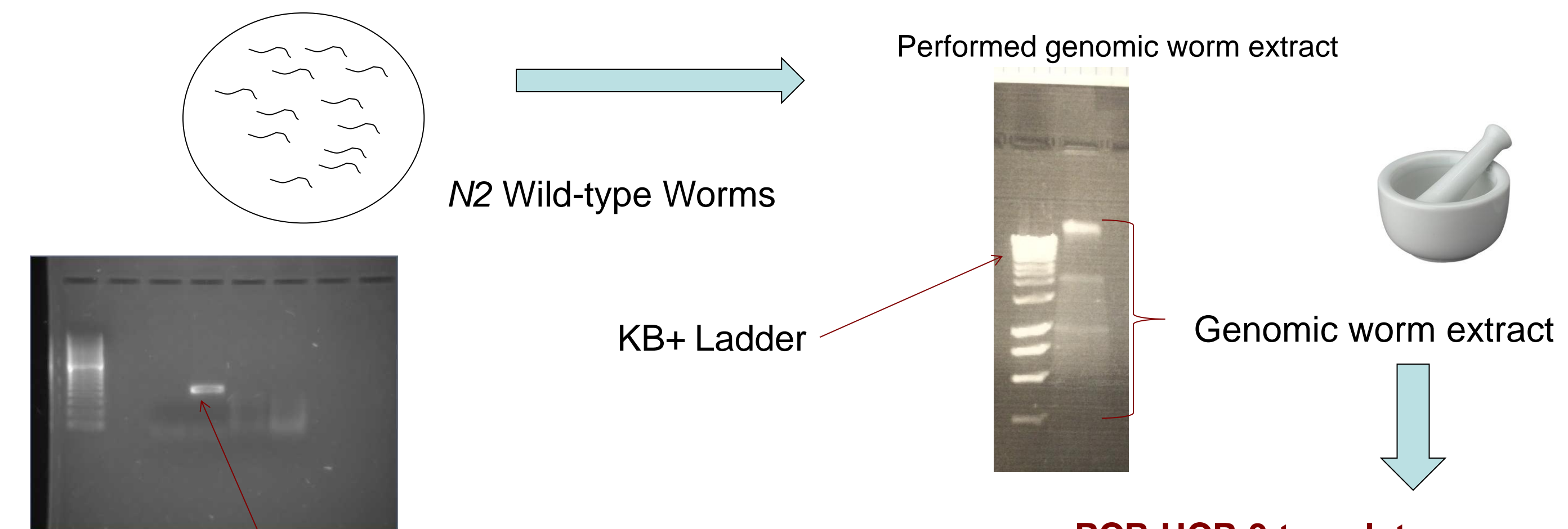
- CPAR-1
- HCP-3



The Setup: Dual Pronged approach to creating and prepping dsRNA for RNAi feeding & injection comparison

RNAi Feeding – Bacteria containing the RNAi plasmid is fed to the worms and when digested will incorporate into the DNA.

RNAi Injection – The RNAi plasmid will be injected directly into the worm gonad causing it to incorporate directly into the DNA



PCR HCP-3 template

Primers used to PCR HCP-3 segment

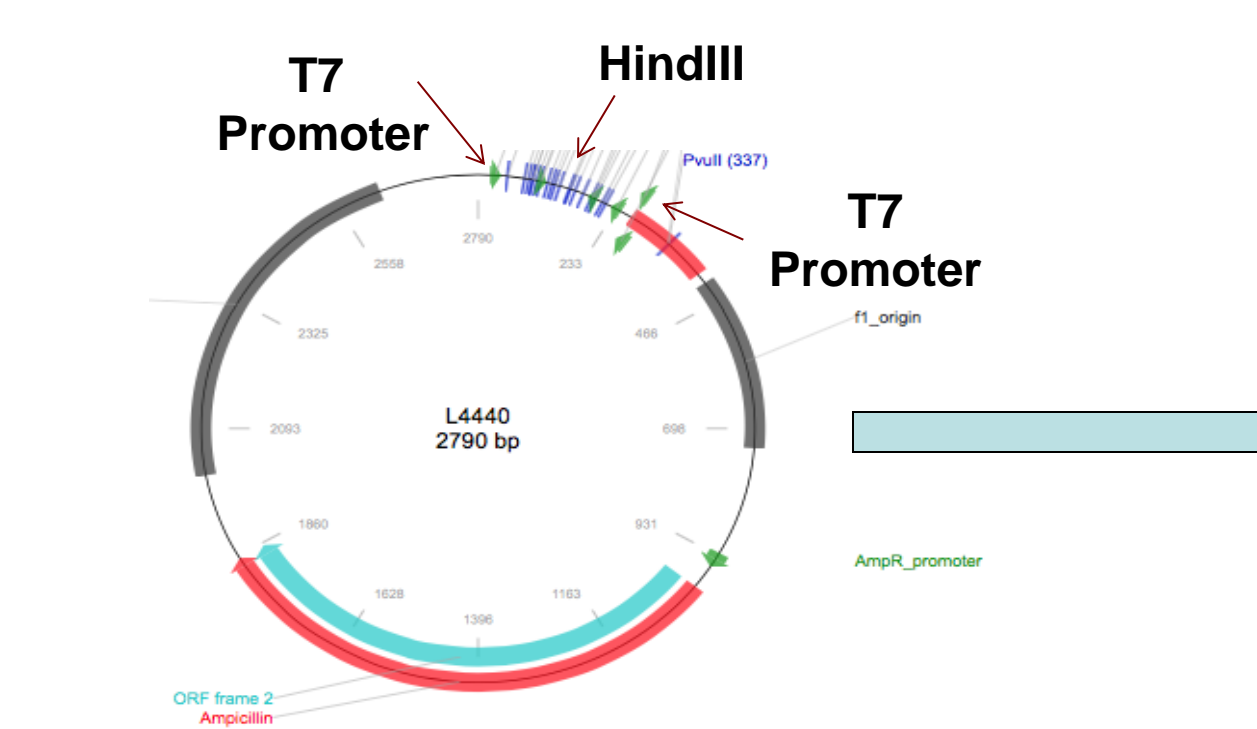
HindIII ends/HCP-3 Forward Primer
GCGGGAAGCTTGGCGGATGACACCCCAATTAT
HindIII ends/HCP-3 Reverse Primer
CGCGCAAGCTTCCTGGGAGTAATCGACAAG

PCR HCP-3 template

Primers used to PCR HCP-3 segment

T7 ends/HCP-3 Forward Primer
TAATACGACTCACTATACGGGCGGTGGAGTAATCGACAAG
T7 ends/HCP-3 Reverse Primer
TAATACGACTCACTATACGGGCGGTGGAGTAATCGACAAG

HCP-3 Gene Sequence



PCR was used to amplify the genomic HCP-3 nucleotide sequence creating the L4440 plasmid

- plasmid will be transformed into the HT115 (DE3) strain of *E. coli*
- will later be fed to the worms
- plasmid contains T7 promoter
- allows for dsRNA to be created *in vitro*

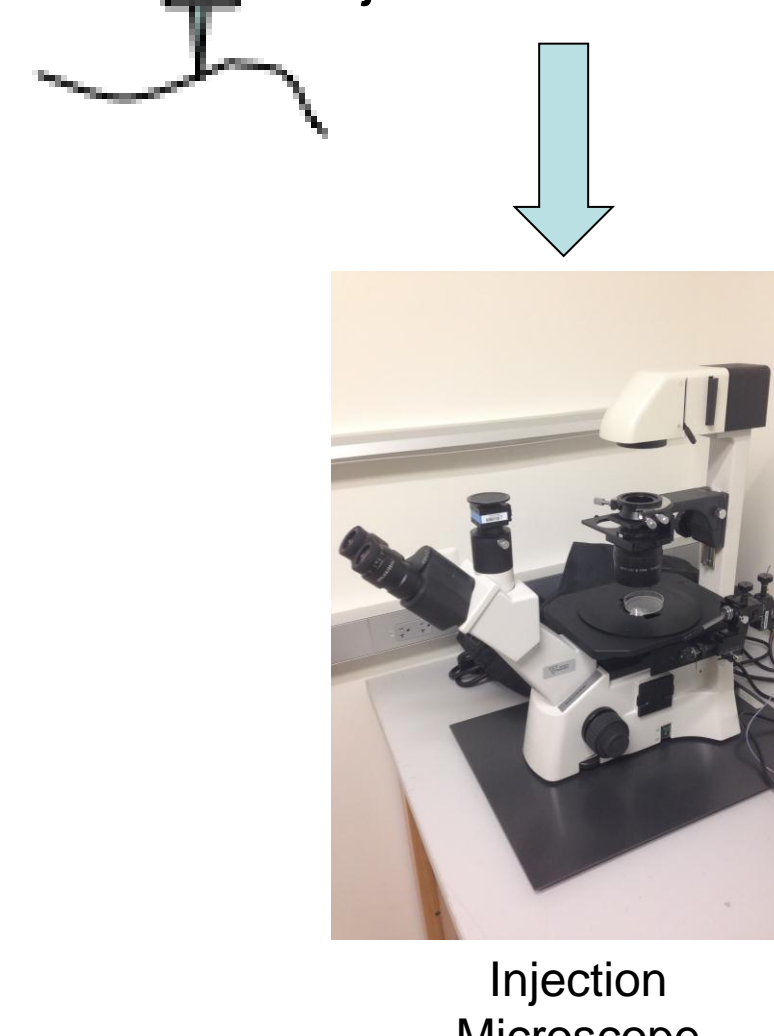
Transform into HT115 (DE3) bacteria

Bacteria containing the dsRNA expression plasmid (pJM13) is fed to the worms and its efficacy can be analyzed

- Confirm presence of pJM13 plasmid
- Grow up and seed plates with HT115 (DE3) for feeding RNAi

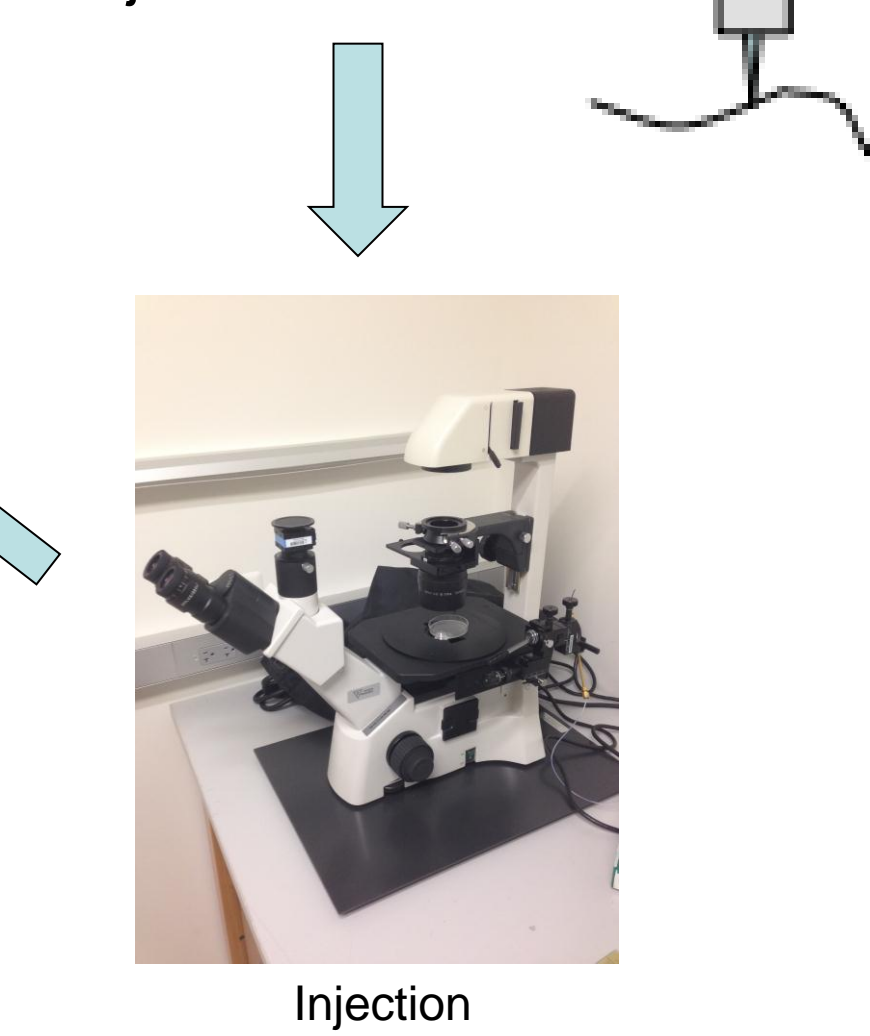
Create control dsRNA *in vitro* using Maxiscript T7 *in vitro* Transcription kit

Inject into L4 worms



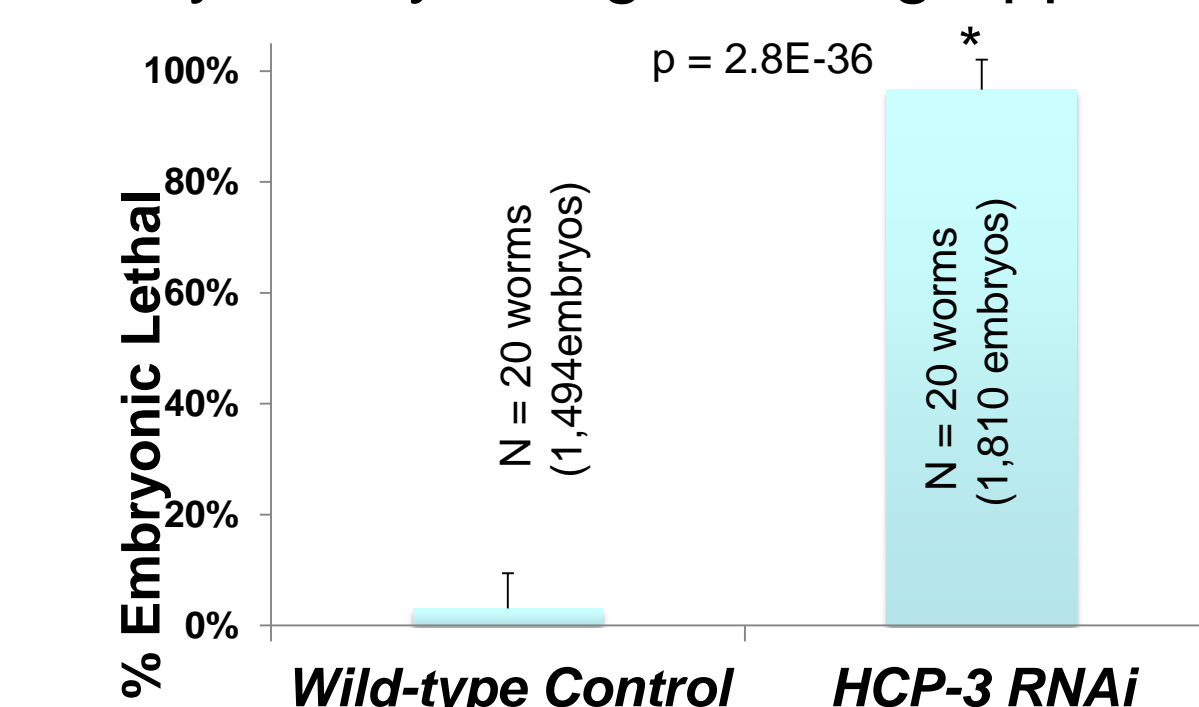
Create dsRNA *in vitro* using Maxiscript T7 *in vitro* Transcription kit

Inject into L4 worms



Incubate for 48 hours
Analyze Control dsRNA injected vs. Experimental dsRNA injected

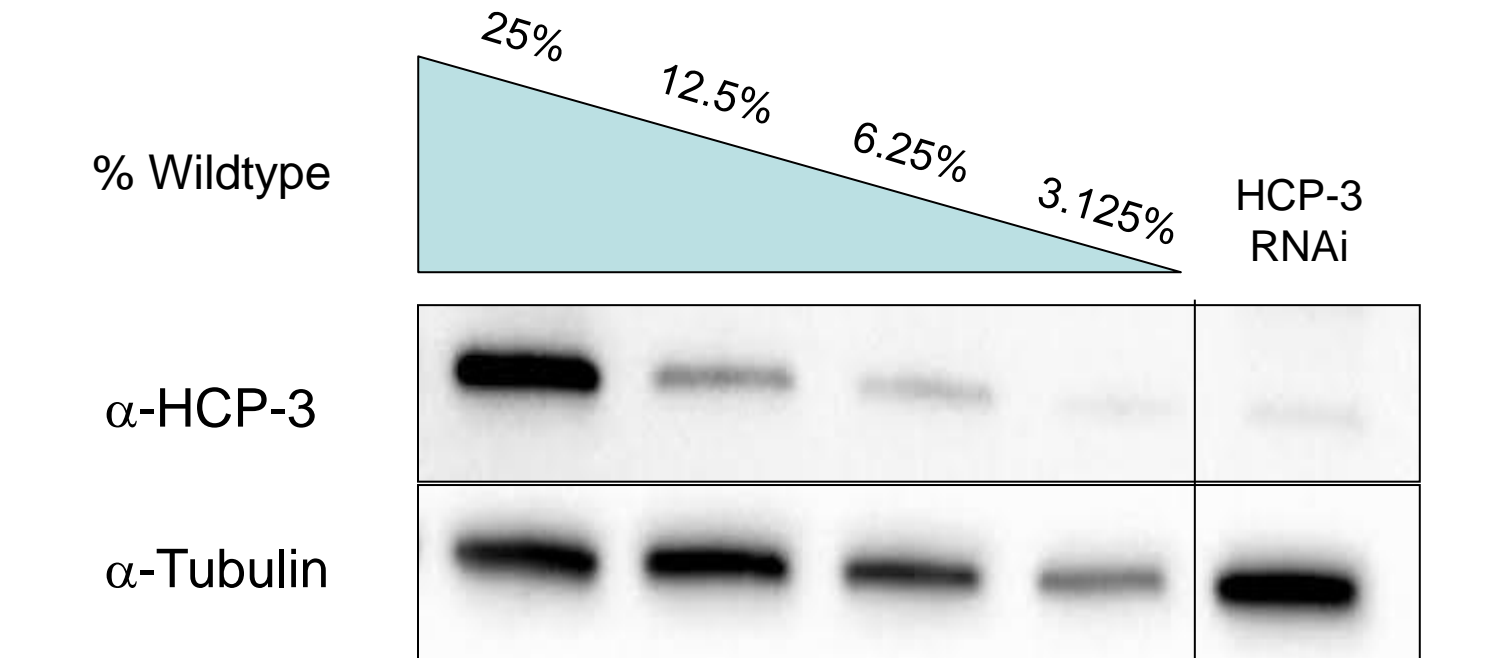
Embryonic Lethality Assay using Feeding Approach



$$\text{Embryonic Lethality} = \frac{\# \text{ of inviable embryos}}{\# \text{ inviable and live worms}}$$

Western Blot

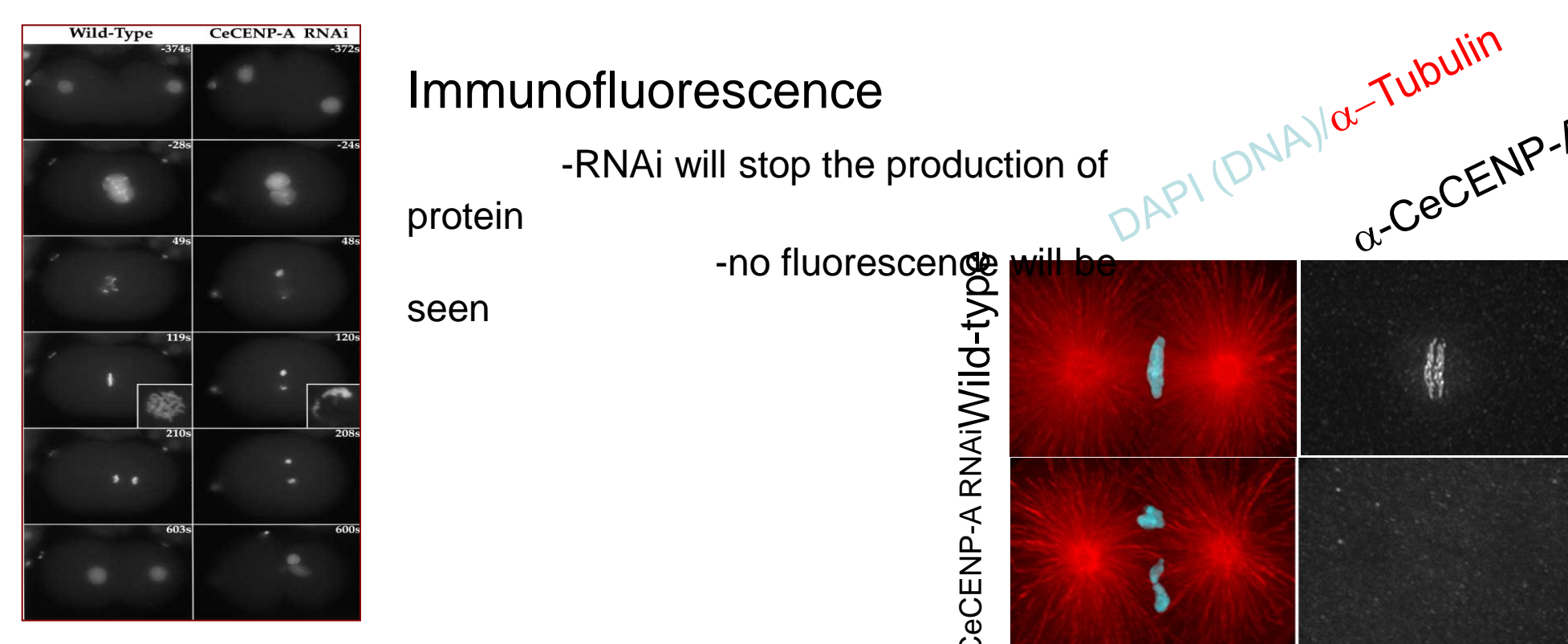
- 1) 100 adult worms from RNAi feeding plate, 100 from control feeding plate to separate M9 tubes
- 2) Wash 3 times, spin, and remove supernatant
- 3) Remove liquid to 50uL and add 50uL 2xSB
- 4) Both tubes were sonicated for 20 minutes
- 5) Serial dilutions for control sample → 22uL of 1xSB in 5 tubes and 22uL from previous tube making each tube half as dilute as the prior tube
- 6) Ran SDS-Page



Future Directions

Live Imaging

- Wild Type goes through the steps of Mitosis naturally and without any issues creating viable offspring
- Cell subjected to CeCENP-A RNAi through RNAi feeding or injection cannot complete Mitosis and dies



References

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Acknowledgements

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